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RESEARCH ARTICLE

COMPARISON OF ANTIBIOTIC SENSITIVITY OF MRSA WITH MSSA AMONG STAPHYLOCOCCUS AUREUS ISOLATES FROM PATIENTS IN THE **48 MILITARY HOSPITAL IN SANA'A CITY, YEMEN**

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ABSTRACT

Background and objectives: A number of infectious disorders can be opportunistically brought on by Staphylococcus aureus (S. aureus), which colonizes human skin and mucous membranes. Methicillin (MRSA) resistance is a frequent occurrence, as are resistances to a number of clinically useful antibiotics. Though MRSA affects healthcare systems and society all throughout the world, it is most severe in underdeveloped countries. The Clinical and Laboratory Standards Institute (CLSI) advises that cumulative antibiotic data for S. aureus be analyzed and reported on an annual basis to help clinicians choose the best preliminary empirical antimicrobial therapy. The most recent report from our center on this subject, however, was more than 7 years ago.

Subjects and methods: Well-proven S. aureus data were gathered from inpatient and outpatient clinical samples at the 48th Military Hospital, Sana'a, Yemen, from January 1, 2022, through December 2022, using a retrospective cross-sectional design. Using Kirby-Bauer disk diffusion, antimicrobial susceptibility testing (AST) was carried out. Calculations were made on the rate of antibiotic resistance between MRSA and MSSA as well as the correlation between MRSA.

Results: Among the 265 unique isolates, the overall prevalence of MRSA was 37.4%. Inpatients had a greater risk factor for MRSA with an OR of 2.7 (p < 0.001). A risk factor was also found with the catheter sample, and devices with an OR=3.7 (p=0.003). Methicillin resistance was predictive of resistance to most antibiotics. Zero resistance rate to linezolid, and vancomycin was observed for the MRSA and MSSA strains. The prevalence of multidrug resistant (MDR) isolates was 60.4%. Significantly higher in MRSA (68.7%) versus 55.4% for MSSA.

Conclusion: This study's MRSA prevalence was higher than that of earlier research from the same hospital; it is a progressive issue and much below the desired rates. Additionally, there was notable resistance to erythromycin, imipenem, and clindamycin. Vancomycin and linezolid are currently the top two options for the empiric treatment of MRSA. In order to stop the emergence of MDR species, it is suggested against giving newer antibacterial medications while the older ones are still effective. Keywords: Antibiotic patterns; multidrug resistant (MDR); MRSA, MSSA, S. aureus, Sana'a, Yemen.

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INTRODUCTION

One of the most prevalent colonizers and a source of several illnesses is S. $aureus^1$. It was found that S. aureus was the second leading pathogen of antimicrobial resistance-related deaths in 2019^2 . It is known that S. aureus isolates became resistant to

penicillin within one to two years of its introduction, methicillin less than a year after its use³, and vancomycin after about 40 years⁴ since its introduction into clinical use. Even while one of them may appear to be effective in vitro, clinical resistance against all other antibiotics in the same class usually results from the mechanism of resistance, which alters the target of the

antibiotic⁵. Staphylococcus bacteria make penicillinase, a specific form of beta-lactamase that cleaves the lactam ring of the penicillin molecule and renders the antibiotic useless. Methicillin, oxacillin, cloxacillin, dicloxacillin, nafcillin, and flucloxacillin are examples of antibiotics with the ability to withstand degradation by staphylococcal penicillinase⁶. At the same time, multidrug resistance (MDR) may coexist against different classes through different mechanisms as well. Methicillin resistance in Staphylococcus aureus (MRSA) itself can be seen as another definition of multidrug resistance⁷. It is associated with several epidemiological features⁸ and may indicate increased resistance against other agents (e.g., clindamycin)⁹. Of course, the use of antibiotics creates selective pressure for MRSA and other resistant isolates, but in developing nations, improper use of antibiotics for common diseases may also contribute to increased resistance. Meanwhile, the high frequency of MRSA in developed countries may be the result of improper use or over-the-counter antibiotics.

Although given to treat a specific disease in a specific patient, antibiotics, unlike other drugs, have effects that extend to much more than just the patient. Antibiotics and antibiotic-resistant bacteria seep into the drinking water supply after use by humans, agricultural animals, and pets, as well as after wastewater treatment. This is true even when prescribed and used correctly. In addition, they frequently contaminate meat and poultry intended for human consumption that are domestically produced or imported, and serve as direct causes of disease or colonization in humans¹⁰. In the current era, due to the recent development and clinical approval of new, powerful antibiotics as new potent antibiotics, using anti-staphylococcal agents sparingly, trying older agents with a narrow/targeted spectrum at the first lines by an appropriate dose and duration, hesitating to prescribe antibiotics in cases where there is no evidence-proven indication, and, if the circumstances allow, waiting for the antibiogram results are all important strategies to follow. Additionally, due to the quick establishment of resistance mutations, rifampin (RIF) or fluoroquino-lones (FQ) monotherapy of S. aureus infections should be avoided¹¹. Another promising finding is the "seesaw effect," which shows increased beta-lactam activity when antibiotics targeting glyco- and/or lipo-peptides are less effective 12 .

For patients for whom there are no yet available microbiological test results to target treatment, the analysis and presentation of cumulative antibiogram reports should occur at least once per year, according to the Clinical and Laboratory Standards Institute (CLSI) $M39^{13}$. The focus of the current investigation is the fact that it has been more than 8 years since the 48 Military Hospital in Sana'a, Yemen had reported *S. aureus*.

SUBJECTS AND METHODS

At the 48 Military hospital, a tertiary referral care facility in Sana'a, Yemen, this cross-sectional retrospective study was carried out. Clinical samples of

specimen types were collected from various hospitalized in-patients and patients attending the outer clinic of the hospital from 1st January 2022 to December 2022. Sample types were considered as follows; wound secretions, blood, respiratory secretion and sputum, abscess, tissue, bone and intra-articular fluid, urine, pleural, peritoneal, and pericardial fluids; catheters and devices and others. Medical records were used to gather information about S. aureus isolates. The first isolate was examined for each patient during a one-year period, regardless of the body site from which the sample was taken or the antimicrobial susceptibility pattern, in accordance with CLSI M39 criteria¹ Furthermore, isolates with missing data were disregarded.

S. aureus identification: In this study, we used phenotypic approaches to identify *S. aureus* isolates and assess their antibiotic susceptibility (AST). To accomplish this, each specimen was examined using a variety of identification techniques, such as Gramstained smear light microscopy, observation of colony morphology and growth patterns on different media, such as deoxyribonuclease agar and mannitol salt agar, and manual biochemical reactions, such as catalase and coagulase tests.

Antibiotic sensitivity: The modified Kirby-Bauer disc diffusion method was used to assess the antibiotic sensitivity of bacterial isolates on Mueller-Hinton agar. The inhibitory zone diameter was interpreted in accordance with the 2017 Clinical Laboratory Standards Institute guidance¹⁵.

Detection of MRSA: Cefoxitin disc diffusion was used to detect MRSA strains as recommended by CLSI to detect methicillin resistance^{15,16}. Cefoxitin can be utilized to screen diverse MRSA populations since it is a more effective inducer of mec-A gene expression than oxacillin or methicillin.

MDR determination: MDR was defined as no susceptibility to ≥ 1 agent in ≥ 3 antimicrobial categories. *Beta-lactamse* was excluded as an antimicrobial category in order to evaluate frequencies of MDR between isolates of MSSA and MRSA. The antibiotic susceptibility (or resistance) pattern in this study identifies the antibiotics to which an isolate is also susceptible (or resistant).

Statistical analysis: Data analysis was done using the Epi Info statistical program, version 6 (CDC, Atlanta, USA). The association of MRSA with baseline characteristics of clinical samples received for *S. aureus* were determine by calculating *OR*, 95% *CI*, X^2 and *p* value. Different antibiotic resistance patterns and their frequency were calculated and difference rate and significance of resistant to different antibiotics were calculated. The significance of the observed difference between groups was assessed using the *Chi* squared test with a threshold *p*-value of 0.05.

RESULTS

The results of the study are illustrated in three tables. Table 1 shows the association of MRSA with baseline characteristics of clinical specimens received of *S. aureus* at the 48^{th} Military Hospital in Sana'a City for

the year 2022. There was an association between MRSA and the inpatient group where the risk factor associated with contracting inpatient MRSA equals 2.7, CI=1.6 -4.4 , p<0.0001. Also, MRSA was associated with abscess, tissue, bone and intra-articular fluid samples with the risk factor associated with MRSA being 1.9, p=0.05. There was an association of MRSA with catheters, devices samples where the risk factor associated with MRSA was 3.7, p=0.003. There was no association with other factors. MRSA isolates were resistant against erythromycin (ERY), clindamycin

(CLI), ciprofloxacin (CIP), and levofloxacin (LVX) by >80%.

All MSSA isolates had resistance rates <50% against each of the tested antibiotics. When reporting the most common antibiotic styles in Table 2, we included nitrofurantoin (NIT) even though it is mainly used for urinary tract infections. Overall, the MDR was 60.4% and was significantly different (*p*-value 0.03) between the MRSA (68.7%) and MSSA (55.4%) isolates with 13.3% difference; and beta-lactams were omitted from the definition of drug resistance (Table 3).

Type of specimens	MSSA	MRSA	OR	CI		$X^2 P$
	n=166 N (%)	N=99 N (%)				
Sex						
Male n=147	98 (66.7)	49 (33.3)	0.68	0.4-1.1	2.2	0.13
Female n=118	68 (57.6)	50 (42.4)	1.4	0.8-2.4	2.2	0.13
Hospitalized						
Yes n= 123	62 (50.4)	61 (49.6)	2.7	1.6-4.4	14.6	< 0.001
No n=142	104 (73.2)	38 (26.8)	0.37	0.2-0.6	14.6	< 0.001
Blood n=23	13 (56.5)	10 (43.5)	1.3	0.59-3.1	0.4	0.52
Wound secretions n=45	30 (66.7)	15 (33.3)	0.8	0.4-1.5	0.37	0.54
Respiratory secretions and	24 (58.5)	16 (41.5)	1.1	0.7-2.2	0.14	0.7
sputum n=41						
Abscess, tissue, bone,	22 (50)	22 (50)	1.9	1.01-3.5	3.6	0.05
intra-articular fluid n=44						
Urine n=25	17 (68)	8 (2)	0.77	0.3-1.8	0.33	0.56
Pleural, peritoneal, and	29 (74.4)	10 (25.6)	0.53	0.24-1.1	2.6	0.1
pericardial fluids n=39						
Catheters and devices =21	14 (66.7)	7 (33.3)	3.7	1.4-9.6	8.3	0.003
Others n=27	16 (59.3)	11 (40.7)	1.2	0.5-2.6	0.14	0.7

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Total	MSSA	MRSA	%	р
S. aureus n=265	n=166	n=99	Difference	
N (R %)	N (R %)	N (R %)		
98 (37)	34 (20.5)	64 (64.6)	43.5	< 0.0001
149 (56.2)	57 (34.3)	92 (93)	58.7	< 0.0001
156 (58.9)	75 (45.2)	81 (81.8)	36.6	< 0.0001
65 (24.5)	40 (24.1)	25 (25.3)	1.2	0.8
122 (46)	55 (33.1)	67 (67.7)	34.6	0.0007
65 (24.5)	5 (3.01)	60 (61)	57.9	< 0.0001
45 (17)	3 (1.8)	42 (42.4)	40.6	< 0.0001
99 (37.4)	0 (0.0)	99 (100)	100	< 0.0001
123 (46.4)	34 (20.5)	89 (89.9)	69.4	< 0.0001
100 (37.7)	53 (31.9)	47 (47.5)	15.6	0.01
116 (43.8)	45 (27.1)	71 (71.7)	44.6	< 0.0001
102 (38.5)	3 (1.8)	99 (100)	98.2	< 0.0001
65 (24.5)	8 (4.8)	57 (57.6)	52.8	< 0.0001
10 (3.8)	0 (0.0)	10 (10.1)	10.1	< 0.0001
0 (0)	0 (0.0)	0 (0.0)	0	-
0 (0)	0 (0.0)	0 (0.0)	0	-
	S. aureus n=265 N (R %) 98 (37) 149 (56.2) 156 (58.9) 65 (24.5) 122 (46) 65 (24.5) 45 (17) 99 (37.4) 123 (46.4) 100 (37.7) 116 (43.8) 102 (38.5) 65 (24.5) 100 (37.7) 116 (43.8) 102 (38.5) 65 (24.5) 10 (3.8) 0 (0)	S. aureus n=265 N (R %)n=166 N (R %)98 (37)34 (20.5)149 (56.2)57 (34.3)156 (58.9)75 (45.2)65 (24.5)40 (24.1)122 (46)55 (33.1)65 (24.5)5 (3.01)45 (17)3 (1.8)99 (37.4)0 (0.0)123 (46.4)34 (20.5)100 (37.7)53 (31.9)116 (43.8)45 (27.1)102 (38.5)3 (1.8)65 (24.5)8 (4.8)10 (3.8)0 (0.0)0 (0)0 (0.0)	S. aureus n=265n=166n=99N (R %)N (R %)N (R %)98 (37)34 (20.5)64 (64.6)149 (56.2)57 (34.3)92 (93)156 (58.9)75 (45.2)81 (81.8)65 (24.5)40 (24.1)25 (25.3)122 (46)55 (33.1)67 (67.7)65 (24.5)5 (3.01)60 (61)45 (17)3 (1.8)42 (42.4)99 (37.4)0 (0.0)99 (100)123 (46.4)34 (20.5)89 (89.9)100 (37.7)53 (31.9)47 (47.5)116 (43.8)45 (27.1)71 (71.7)102 (38.5)3 (1.8)99 (100)65 (24.5)8 (4.8)57 (57.6)10 (3.8)0 (0.0)10 (10.1)0 (0)0 (0.0)0 (0.0)	S. aureus n=265 N (R %)n=166 N (R %)n=99 N (R %)Difference Difference98 (37)34 (20.5)64 (64.6)43.5149 (56.2)57 (34.3)92 (93)58.7156 (58.9)75 (45.2)81 (81.8)36.665 (24.5)40 (24.1)25 (25.3)1.2122 (46)55 (33.1)67 (67.7)34.665 (24.5)5 (30.1)60 (61)57.945 (17)3 (1.8)42 (42.4)40.699 (37.4)0 (0.0)99 (100)100123 (46.4)34 (20.5)89 (89.9)69.4100 (37.7)53 (31.9)47 (47.5)15.6116 (43.8)45 (27.1)71 (71.7)44.6102 (38.5)3 (1.8)99 (100)98.265 (24.5)8 (4.8)57 (57.6)52.810 (3.8)0 (0.0)10 (10.1)10.10 (0)0 (0.0)0 (0.0)0

DISCUSSION

The goal of this investigation was to identify the cumulative antibiotics pattern (Table 2) for *S. aureus* isolates in the 48th Military Hospital in order to be included in antibiotic stewardship programs, as advised by CLSI M39¹³. After analyzing 265 *S. aureus* isolates from clinical samples, the overall prevalence of MRSA isolates was 37.4%. Over 80% resistance rates against Erythromycin, Clindamycin, Ciprofloxacin and

imipenem were seen among MRSA isolates which is of concern because Clindamycin and imipenem are two of the most commonly prescribed antibiotics empirically. No resistance was discovered with linezolid (LZD), and vancomycin (VAN). Although this was the case, in hospital settings, MRSA remained a rare occurrence until the 1990s, at which point its prevalence in hospitals surged and it is now endemic¹⁷. Now, MRSA infecting humans and causing a number of infections, including skin and soft tissue infection (SSTI), pneumonia, and sepsis, it can also infect animals, causing livestock-associated MRSA (LA-MRSA) sickness¹⁸. In the current study, the prevalence of MRSA was lower than previous reports by Al-Safani *et al.*, in the same center⁶ (19%), Khalili *et al.*,¹⁹ and Mehrez *et al.*,²⁰ in Iran, as well as less than that reported in Yemen by Al-Akwa *et al.*,²¹ (23.5%). Alyahawi *et al.*, (17.6%)²² but similar to that recently reported by Qodrati *et al.*,²³ in Iran (37.5%). When a comparison of isolates causing invasive infection from 29 European countries in 2018 is made, the current result will be placed after Cyprus, Romania and

Portugal, in fourth place and these countries are among the countries with the most prevalence of MRSA. Additionally, the general rate in Europe is 19.3% in the same report²⁴. In the present study, the overall prevalence of MDR isolates was 60.4%, exceeding the rates determined in Addis Abeba by Dilnessa and Bitew⁹ and Iran by Qodrati *et al.*,²³ (48.5%). Additionally, it was significantly lower than what Kim *et al.*,²⁵ studied with a tailored definition (97.7%) and significantly higher than what Wiliamson *et al.*,²⁶ reported from New Zealand (6%).

Table 3: Prevalence of MDR degree among S. aureus of MSSA strains comparing with MRSA strains isolated
from clinical specimens.

Antimicrobial class used to define MDR	Degree	Total S. <i>aureus</i>	MSSA N=166	MRSA N=99	Difference %	р
define wiDk		S. aureus N=265, N (%)	N=100 N (%)	N=99 N (%)	70	
1-Glycopeptide (Vancomycin)	R0	50 (18.9)	37 (22.3)	13 (13.1)	9.2	0.64
2-Aminoglycosides (Gentamicin)	R1	10 (3.8)	2 (1.2)	8 (8)	6.8	0.004
4-Quinolone (Ciprofloxacin)	R2	45 (17)	35 (21.1)	10 (10)	9.2	0.09
5-Sulfonamides (Cotrimoxazole)	R3	20 (7.5)	17 (10.2)	3 (3)	7.2	0.03
6-Oxazolidinones (Linezolid)	R4	65 (24.5)	35 (21.1)	30 (30.3)	9.2	0.92
7-Macrolides (Erthromycin)	R5	35 (13.2)	19 (11.4)	16 (16.2)	4.8	0.26
Total MDR=160 (60.4%)	R6	25 (9.4)	12 (7.2)	13 (13.1)	5.9	0.11
	R7	15 (5.7)	9 (5.4)	6 (6)	0.6	0.83
	MDR	160 (60.4)	92 (55.4)	68 (68.7)	13.3	0.03

R0: Sensitive against all selected antibiotic class; R1: Resistant to at least one antibiotic class; R2: Resistant to two antibiotic class; R3: Resistant to four antibiotic class; R5: Resistant to five antibiotic class; R6: Resistant to six antibiotic class; R7: Resistant to all seven antibiotic class; MDR: Resistant to at least three antibiotic class.

Inpatients had a higher chance of being infected with MRSA isolates with an OR of 2.7 (p<0.001) compared to outpatients. The above result is reasonable; Most infections occur in the community, which are linked to organisms that are least resistant. The infections that appear in hospital acquired infections are caused by pathogens that are more resistant to antibiotics and that also increase the overall resistance rate of *S. aureus*. MRSA rates were significantly different between sample types; Abscess, tissue, bone, and intra-articular fluid had a higher chance of with MRSA with *OR*=1.9 (p=0.057) and catheters and devices with *OR*=3.7 (p=0.003). This was contrary to what Mehraz *et al.*,²⁰ found Waitayangkun *et al.*,⁸ or Dilnessa and Bitew⁹.

Methicillin resistance considerably increased the resistance status against the majority of antibiotics. It was usual to anticipate 100% resistance to other betalactams (imipenem). Levofloxacin, clindamycin, tetracycline, erythromycin, gentamicin, rifampicin, and trimethoprim/sulfamethoxazole are likely to be ineffective against the MRSA isolate, and no difference was seen between MSSA and MRSA when tested with chloramphenicol, nitrofurantoin, linezolid (LZD), and vancomycin. These findings concurred with those of earlier studies^{9,23,27,28}. The prevalence of clindamycin resistance in the current investigation, which also included inducible clindamycin resistance, was 56.2% overall and 93% for MRSA isolates. When there is a low resistance rate (e.g., 10%), the Infectious Diseases Society of America (IDSA) guidelines advice²⁸ empirically treating skin and soft tissue MRSA infections with clindamycin. As a result, the current findings do not support the use of clindamycin in Sana'a, Yemen. Although high-level vancomycinresistant aureus isolates from Yemen were reported before $(40\%)^6$, the prevalence rate of vancomycin intermediate S. aureus was reported by Al-Shami et al., $(1.4\%)^{30}$. In Yemen, the results of the current study appear promising with the result of Al-Safani et al., (40%) and also according to Al-Shami *et al.*, $(1.4\%)^{30}$. Out of the three newer antibiotics, LZD has the highest clinical availability and is the only oral option with a 100% susceptibility rate in current study. Similar rates were seen in previous studies^{31,32}, but Baddour et al.,⁵ with a 4.1% resistance rate demonstrated that the establishment of LZD-resistance has already started and is a progressive trend over time. Although these drugs are beneficial additions to Yemen's antimicrobial options, their usage should be restricted to patients who actually need them in order to delay the evolution of antibiotic resistance in Yemen and globally.

The rate of MRSA resistance against gentamicin in the current study was 61% versus 3.01% for MSSA. Streptomycin, kanamycin, and gentamicin were once effective against *staphylococcal* infections until strains developed resistance against them. Aminoglycosides work by interfering with ribosomal RNA of the 30S subunit of the ribosome through amine and/or hydroxyl interactions³³. Aminoglycoside-modifying enzymes, ribosomal mutagenesis, and active efflux of the drug out of bacteria are the three primary mechanisms of aminoglycoside resistance that are currently and widely acknowledged⁶. By covalently joining a phosphate moiety, a nucleotide, or an acetyl to either the primary amine or alcohol functional group (or both groups) of the antibiotic, aminoglycoside-modifying enzymes

render aminoglycosides inactive. This alters the antibiotic's charge or sterically inhibits it, lowering its affinity for attaching to ribosomes. Aminoglycoside adenylyltransferase 4' IA (ANT(4')IA) is the aminoglycoside modifying enzyme that has been most studied in S. aureus. X-ray crystallography has been used to identify this enzyme³⁴. Many aminoglycosides, notably kamamycin and gentamicin, have a 4' hydroxyl group that the enzyme is able to link an adenyl moiety to. The current investigation found a 0.0% MRSA resistance rate to glycopeptides and vancomycin. The Tn1546 transposon, which was discovered in a plasmid in enterococci, is the source of the van-A gene, which codes for an enzyme that creates an alternate peptidoglycan that vancomycin will not bind to³⁵. This alternative peptidoglycan is what mediates glycopeptide resistance.

Non-lactam antibiotics, such as clindamycin (a lincosamine) and trimethoprim/sulfamethoxazole, are frequently used to treat MRSA infections in both the hospital and the community. Due to linezolid's accessibility as an oral medication, resistance to these antibiotics has also prompted the adoption of new, broad-spectrum anti-Gram-positive antibiotics. Glycopeptide antibiotics (vancomycin and teicoplanin) are presently the first-line treatment for significant invasive infections caused by MRSA. These antibiotics have a variety of drawbacks, including the requirement for intravenous administration (no oral preparation is available), toxicity, and the requirement to routinely check medication levels through blood testing. Additionally, glycopeptide antibiotics do not penetrate infected tissues very well (this is especially problematic for endocarditis and infections of the brain and meninges). Methicillin-sensitive S. aureus (MSSA) should not be treated with glycopeptides because the results are subpar³⁶.

Due to the high rate of penicillin resistance and the potential for MRSA to develop vancomycin resistance, the U.S. Centers for Disease Control and Prevention have produced guidelines for the appropriate use of vancomycin. In situations where the prevalence of MRSA infections is known to be high, the attending physician may choose to use a glycopeptide antibiotic until the identity of the infecting organism is established. Once a MSSA strain of *S. aureus* has been identified as the cause of the infection, the appropriate course of treatment can be altered to flucloxacillin or even penicillin²³.

Limitations of the study

The shortcomings of the study were as follows. First of all, because the data came from a single center, it was unable to accurately identify multidrug resistance in each MRSA isolate for each hospital in Sana'a. Second, no molecular research has been performed on these isolates to support the findings.

CONCLUSION

Overall, this study found that MRSA frequency was alarmingly high compared to earlier studies conducted at the same hospital eight years earlier. Additionally, there appeared to be an unacceptable level of resistance to popular alternative antibiotics as clindamycin and trimethoprim-sulfamethoxazole, while a *S. aureus* infection is detected, it may be more fair to empirically begin with first-generation cephalosporins rather than clindamycin, and the natural course and response to therapy should be further taken into account while escalating the antimicrobial regimen. Vancomycin is now the gold standard for treating MRSA infections due to its low resistance rate and availability in comparison to newer drugs that are more expensive and have more adverse effects. The sole oral medication that has gained popularity for treating MRSA infections is linezolid, although it is best to save these medications for last-resort use if the rate of vancomycin resistance rises significantly in the future.

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CONFLICT OF INTEREST

Regarding this project, there is no conflict of interest.

AUTHOR CONTRIBUTIONS

The research was approved by all authors after it had been revised, written, and the data had been assessed.

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